JRRENT STATE OF ANTIMICROBIAL RESISTANCE OF *S. PNEUMONIAE* IN RUSSIA: RESULTS OF PROSPECTIVE R.S. Kozlov^{1*}, O.V.Sivaja¹, O.I. Kretchikova¹, E.D. Agapova², L.I. Ahmetova³, M.E. Furletova⁴, L.V. Gudkova⁵, E.N. Gugutsidse⁶, R.R. Egorova⁷, V.N. Ilyina⁸, L.K. Katosova⁹, F.K. Manerov¹⁰, N.E. Marusina¹¹, I.G. Multich¹², G.I. Nechaeva¹³, N.M. Nurtdinova¹⁴, E.A. Ortenberg¹⁵, S. H. Paljutin¹⁶, I.V. Smirnov¹⁷, E.V. Schetinin¹⁸, S.M. Shturmina¹⁹, G.Ya. Tseneva²⁰, L.S. Stratchounski¹ (1000) *Contact information: Dr. Roman S. Kozlov, 214019, Russia, Smolensk, P.O. Box 5, Email: roman@antibiotic.ru

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Abstract

Purpose: To determine the antimicrobial resistance of clinical strains *S. pneumoniae* isolated in adults and children.

Methods: This study was conducted in 20 cities (Chelyabinsk, Ekaterinburg, Irkutsk, Yakutsk, Yaroslavl, Kazan, Kovrov, Krasnodar, Moscow, N. Novgorod, Novosibirsk, Novokuznetsk, Saint Petersburg, Smolensk, Stavropol, Tyumen, Tomsk, Ryazan, Ufa, Voronezh) in different regions of Russia in 2001-2002. Identification of the strains was done on the basis of colony morphology, Gram stain, optochin susceptibility and bile solubility tests. Susceptibility to penicillin G (PEN), amoxicillin (AMO), amoxicillin/clavulanate (AMC), cefotaxime (CTX), cefepime (CFP), imipenem (IMP), erythromycin (ERY), azithromycin (AZI), clarithromycin (CLA), midecamycin (MID), clindamycin (CLI), telithromycin (TEL), levofloxacin (LEV), moxifloxacin (MOX), tetracycline (TET), co-trimoxazole (SXT), chloramphenicol (CHL), vancomycin (VAN) was determined by broth microdilution. Breakpoints were those of NCCLS (2002) except for TEL (≤ 0.5 ; 1-2, \geq 4 mg/L) and MID (\leq 1; 2-4, > 4 mg/L) for susceptible, intermediate resistant and resistant isolates, respectively.

Results: The total of 581 non-duplicate clinical strains of *S. pneumoniae* were included in this study. The susceptibility testing results are presented in the Table 1.

MIC breakpoints MIC₉₀, MIC range, Antimicrobial I/R% mg/L mg/L R S 0.125 8.3/1.9 **≼0.06** 0.12-1 ≥2 0.008-2 PEN ≦2 ≥8 0.06 AMO 0 0.03-2 4 AMC ≦2 ≥8 0.06 0.03-2 0 4 **≤0.5*/1** 1/2 ≥2*/4 0.2/0* 0.03 0.008-1 CTX **≼0.5*/1** ≥2*/4 0.2/0* CFP 1/2 0.125 0.008-2 0.06 0.008-2 IMP 2.6/0.3**≼0.12** 0.25-0.5 ≥1 0.2/8.8 **≼0.25** 0.06 0.016-128 0.5 ERY ≥1 AZI 0.5/8.3 0.03-128 **≼0.5** ≥2 0.125 1 0.5/8.1 0.016-128 **≼0.25** 0.5 ≥1 0.06 CLA 0.5/4.0 0.05-128 0.5 MID ≥4 ≦1 2 0.5 0.2/3.30.03 0.016-128 CLI **≼0.25** ≥1 **≼0.5** 1-2 ≥4 0.03 0.004-0.25 TEL 0 LEV ≤2 ≥8 0.25-2 4 0 1 0.015-0.5 ≥4 0.125 MOX ≼1 2 0 2.4/25.1 0.25-64 ≥8 TET ≤2 16 4 SXT 26.5/5.0 0.06-16 **≼0.5** 1-2 ≥4 2 0.5-16 CHL ≼4 ≥8 0/8.6 4 — VAN ≼1 0 0.5 0.06-1 _ _ * – for non-invasive isolates

Table 1. Non-susceptibility rates, MIC_{90s} and MIC ranges of isolated S. pneumoniae

Conclusions: All *B*-lactams retained high activity against *S. pneumoniae*. High resistance to TET and SXT compromises their usage for the empirical therapy of pneumococcal infections. LEV, MOX, TEL and VAN demonstrated excellent in vitro activity against both penicillin- and macrolide-resistant strains.

S. pneumoniae is one of the most common bacterial pathogens in children and adults causing community-acquired respiratory tract infections (e.g. acute otitis media, sinusitis, pneumonia, meningitis etc.) which are among the most frequent reasons for seeking of medical advice. Currently the prevalence of antibiotic resistance is increasing globally. For the time being, B-lactams, macrolides and fluoroquinolones are recommended as drugs of choice for the variety of pneumococcal infections. Situation with resistance is not uniform and there are substantial differences in patterns of resistance do exist between countries and regions. Thus, regional and local data on resistance are of extreme importance.

To determine the antimicrobial resistance of clinical strains S. pneumoniae isolated in adults and children in different regions of Russia.

This study was conducted in 20 cities (Chelyabinsk, Ekaterinburg, Irkutsk, Yakutsk, Yaroslavl, Kazan, Kovrov, Krasnodar, Moscow, N. Novgorod, Novosibirsk, Novokuznetsk, Saint Petersburg, Smolensk, Stavropol, Tyumen, Tomsk, Ryazan, Ufa, Voronezh) in Russia (Fig. 1). Identification of the strains was done on the basis of colony morphology, Gram strain, optochin susceptibility and bile solubility tests. Susceptibility testing was performed using cationadjusted Mueller-Hinton broth (BBL, USA) with 2-5% lysed horse blood. Microtiter plates were incubated for 24 h at 35°C at ambient air. S. pneumoniae ATCC 49619 was used for quality control. Interpretation of results was done according to NCCLS guidelines (2002).

Introduction

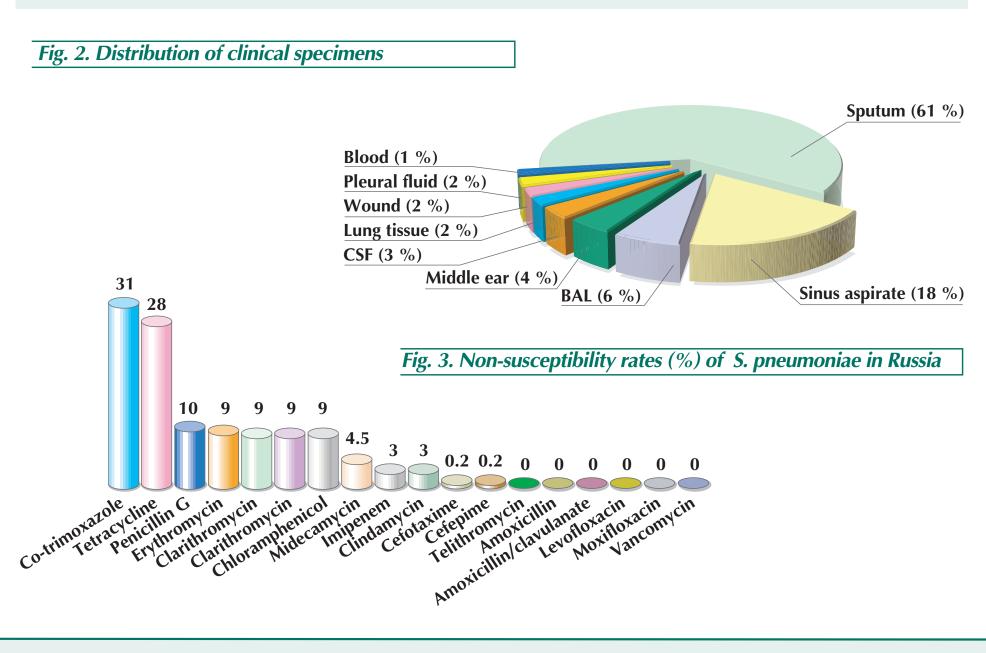
Purpose

Methods



A total of 581 non-duplicate clinical strains of S. pneumoniae isolated from patients of 1 month to 87 years were included in this study. Clinical specimens from which S. pneumoniae have been isolated are presented in Fig. 2. The majority of strains were isolated from respiratory specimens (89%) and 4% – from sterile sources.

The percentages of non-susceptible (intermediately resistant plus resistant) to tested antimicrobials isolates are presented in Fig. 3. The MIC distribution of PEN, ERY, CLI, MID, TET and SXT are presented on Fig. 4-9. The majority 61.5% (32 of 52) of erythromycin-resistant *S. pneumoniae* retained susceptibility to 16-membered macrolides (midecamycin) and clindamycin, indicating M-phenotype of resistance (Fig. 10).



Results

